

Dynabeads® Untouched™ Mouse T Cells

Catalog no. 11413D

Store at 2°C to 8°C

Rev. Date: March 2012 (Rev. 005)

Kit Contents

Kit contents	Volume
Mouse Depletion Dynabeads®	2 × 10 mL
Antibody Mix (for Mouse T cells)	2 mL

Kit capacity
~1 × 10⁹ cells

Mouse Depletion Dynabeads® contains 4 × 10⁸ beads/mL in phosphate buffered saline (PBS), pH 7.4, containing 0.1% bovine serum albumin (BSA) and 0.02% sodium azide as a preservative. The Antibody Mix for mouse T cells contains a mixture of rat monoclonal antibodies in PBS with 0.02% sodium azide.
Caution: Sodium azide may react with lead and copper plumbing to form highly explosive metal azides.

Product Description

This product is intended for isolation of untouched mouse T cells from mouse spleen or lymph node cells by depleting B cells, monocytes/macrophages, NK cells, dendritic cells, erythrocytes, and granulocytes. Isolated untouched mouse T cells are bead- and antibody-free and suitable for use in any downstream application (fig. 1). Add a mixture of monoclonal rat IgG antibodies against the non-T cells to the starting sample to label the unwanted cells.

Wash the cells and add Mouse Depletion Dynabeads® to bind to the antibody labeled cells during a short incubation.

Apply to magnet and transfer the supernatant with the untouched mouse T cells to a new tube and discard the bead-bound cells.

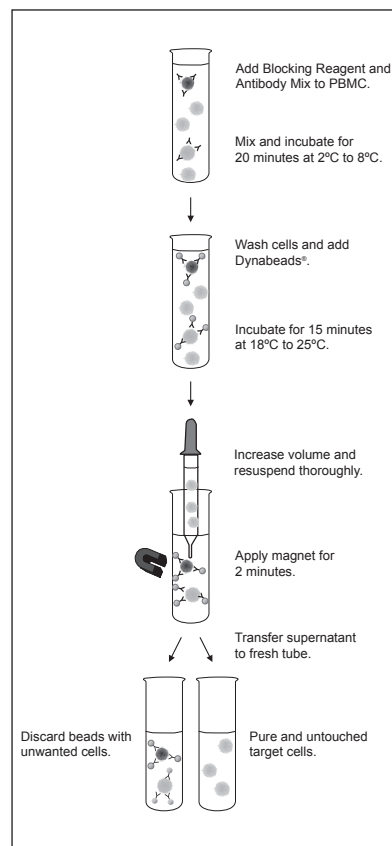


Figure 1: Simple method for isolating untouched mouse T cells.

Downstream Applications

Isolated mouse T cells can be used in any application, (e.g., studies on T cell proliferation, studies on antigen-specific T cells, studies on regulation of T cell cytokine expression, flow cytometry/FACS sorting). Isolated cells can be activated/expanded using Dynabeads® Mouse T-Activator CD3/CD28 (polyclonal activation) or Dynabeads® Mouse T-Activator CD3/CD28/CD137 (antigen-specific activation).

Required Materials

- Magnet (DynaMag™ portfolio). See www.lifetechnologies.com/magnets for recommendations.
- Mixing device with tilting and rotation, e.g. HulaMixer® Sample Mixer.
- Heat inactivated Fetal Bovine Serum (FBS)/Fetal Calf Serum (FCS).
- Isolation Buffer: PBS (Ca²⁺ and Mg²⁺ free) supplemented with 0.1% BSA and 2 mM EDTA.
Note: BSA can be replaced by human serum albumin (HSA) or 2% FBS/FCS. EDTA can be replaced by 0.6% sodium citrate.

General Guidelines

- Visit www.lifetechnologies.com/samplepreparation for recommended sample preparation procedures.
- Use a mixer that provides tilting and rotation of the tubes to ensure that beads do not settle in the tube.
- This product should not be used with the MPC™-1 magnet (Cat. no. 12001D).
- Follow the recommended volumes and incubation times.
- Avoid air bubbles (foaming) during pipetting.
- Keep the buffers cold.

Protocol

Wash the Beads

See Table 1 for volume recommendations.

1. Resuspend the beads in the vial (i.e. vortex for >30 sec, or tilt and rotate for 5 min).
2. Transfer the desired volume of beads to a tube.
3. Add the same volume of Isolation Buffer, or at least 1 mL, and resuspend.
4. Place the tube in a magnet for 1 min and discard the supernatant.
5. Remove the tube from the magnet and resuspend the washed beads in the same volume of Isolation Buffer as the initial volume transferred of beads (step 2).

Prepare Sample

- Prepare spleen or lymph node cells according to “General Guidelines”.
- Resuspend the cells at 1 × 10⁸ cells/mL in Isolation Buffer. The protocol might need to be optimized if the cells are isolated from other sources.

Isolate Untouched Mouse T Cells

This protocol is based on 5×10^7 leucocytes, but it is scalable from $1 \times 10^7 - 1 \times 10^9$ cells, see Table 1.

1. Transfer 500 μ L (5×10^7) leucocytes in Isolation Buffer to a tube.
2. Add 100 μ L heat inactivated FCS/FBS.
3. Add 100 μ L of Antibody Mix.
4. Mix well and incubate for 20 min at 2°C to 8°C.
5. Wash the cells by adding 10 mL Isolation Buffer. Mix well by tilting the tube several times and centrifuge at $350 \times g$ for 8 min at 2°C to 8°C. Discard the supernatant.
6. Resuspend the cells in 4 mL Isolation Buffer.
7. Add 1 mL pre-washed and resuspended Mouse Depletion Dynabeads®.
8. Incubate for 15 min at 18°C to 25°C with gentle tilting and rotation.
9. Add 5 mL Isolation Buffer.
10. Resuspend the bead-bound cells by gently pipetting 5 times using a pipette with a narrow tip opening. Avoid foaming.
11. Place the tube in the magnet for 2 min and transfer the supernatant containing the untouched T cells to a new tube. Discard the beads with the unwanted cells.

Table 1: Volumes for isolation of mouse T cells. This protocol is scalable from 1×10^7 to 3×10^8 leucocytes.

Step	Step description	Volumes per 5×10^7 leucocytes	Volumes per 3×10^8 leucocytes
	Recommended tube	15 mL tubes	50 mL tubes
	Recommended magnet	DynaMag™-15	DynaMag™-50
1	Cell volume	500 μ L	3 mL
2	FCS/FBS	100 μ L	600 μ L
3	Antibody Mix for mouse T cells	100 μ L	600 μ L
5*	Wash cells (Isolation Buffer)	~10 mL	~30 mL
6	Resuspend cells (Isolation Buffer)	4 mL	~24 mL
7**	Mouse Depletion Dynabeads®	1 mL	6 mL
9*	Volume added before magnet	~5 mL	~15 mL

* Adjust the Isolation Buffer volumes to fit to the tube you are using.

** When incubating, tilt and rotate so the cells and beads are kept in the bottom of the tube. Do not perform end-over-end mixing if the volume is small relative to the tube size.

Description of Materials

Mouse Depletion Dynabeads® are uniform, superparamagnetic polystyrene beads (4.5 μ m diameter) coated with a polyclonal sheep anti-rat IgG antibody. The Antibody Mix for mouse T cells contains a mixture of rat monoclonal antibodies against mouse CD45R (B220), CD11b (Mac1), Ter-119, and CD16/CD32.

Related Products

Product	Cat. no.
DynaMag™-5	12303D
DynaMag™-15	12301D
DynaMag™-50	12302D
HulaMixer® Sample Mixer	15920D
Dynabeads® Mouse T-Activator CD3/CD28	11452D
Dynabeads® Mouse T-Activator CD3/CD28/CD137	11454D

[REF] on labels is the symbol for catalog number.

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