

Dynabeads® CD25

Catalog no. 11157D

Store at 2°C to 8°C

Rev. Date: March 2012 (Rev. 003)

Product Contents

Product contents	Volume
Dynabeads® CD25	5 mL

Product capacity

Whole blood: 200 mL MNC: \sim 5 × 10 9 cells

Dynabeads® CD25 contains 4×10^8 beads/mL in phosphate buffered saline (PBS), pH 7.4, with 0.1% bovine serum albumin (BSA) and 0.02% sodium azide as a preservative.

Caution: Sodium azide may react with lead and copper plumbing to form highly explosive metal azides.

Product Description

Isolate or deplete human CD25⁺ T cells directly from whole blood, buffy coat or mononuclear cells (MNC) with Dynabeads® CD25. For rapid and consistent results in protein or gene expression analysis, lyse the CD25⁺ T cells while they are still attached to the beads and directly process for further molecular analysis. The beads are mixed with the cell sample in a tube. The beads bind to the target cells during a short incubation, and then the bead-bound cells are separated by a magnet (fig. 1).

Depletion – Discard the beadbound cells and use the remaining, untouched cells for any application.

Positive isolation – Discard the supernatant and use the beadbound cells for downstream molecular applications.

Downstream Applications

CD25+ cells can be efficiently depleted from a sample. For rapid and consistent results in protein or gene expression analysis, lyse the CD25⁺ T cells while still attached to the beads and directly process for further molecular analysis. For positive isolation for functional studies, cell activation/expansion, or for flow cytometer analysis, the cells need to be released after isolation. For this, we recommend using Dynabeads® FlowComp™ Flexi with your own CD25 antibody (bead-free cells). For isolation of CD4+CD25+ regulatory T cells, see "Related Products".

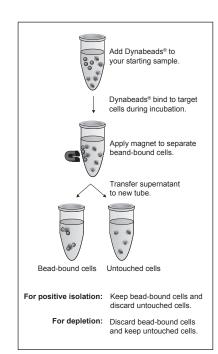


Figure 1: Overview of method

Required Materials

- Magnet (DynaMag[™] portfolio). See www.lifetechnologies.com/magnets for recommendations.
- Mixer allowing tilting and rotation of tubes (e.g. HulaMixer® Sample Mixer).
- Isolation Buffer: Ca²⁺ and Mg²⁺ free PBS supplemented with 0.1% BSA and 2 mM EDTA, pH 7.4.

Note: BSA can be replaced by human serum albumin (HSA) or fetal calf serum (FCS). EDTA can be replaced by sodium citrate.

General Guidelines

- Visit www.lifetechnologies.com/samplepreparation for recommended sample preparation procedures.
- Use a mixer that provides tilting and rotation of the tubes to ensure that beads do not settle in the tube.
- This product should not be used with the MPC[™]-1 magnet (Cat. no. 12001D).
- Avoid air bubbles (foaming) during pipetting.
- Carefully follow the recommended pipetting volumes and incubation times.
- Keep all buffers cold.

Protocol

Approximately 1×10^6 T cells are present/mL human blood, and about 5–10% of these T cells strongly express the CD25 antigen. This protocol describes depletion or positive isolation of CD25⁺ T cells from MNC using Dynabeads® CD25.

Wash the Beads

See Table 1 for volume recommendations.

- 1. Resuspend the beads in the vial (i.e. vortex for >30 sec, or tilt and rotate for 5 min).
- 2. Transfer the desired volume of beads to a tube.
- 3. Add the same volume of Isolation Buffer, or at least 1 mL, and resuspend.
- 4. Place the tube in a magnet for 1 min and discard the supernatant.
- 5. Remove the tube from the magnet and resuspend the washed beads in the same volume of Isolation Buffer as the initial volume of beads (step 2).

Prepare Cells

- Cells can be directly isolated from any sample such as whole blood, bone marrow, MNC suspensions or tissue digests.
- Prepare MNC to 2.5×10^7 cells/mL in Isolation Buffer.
- See "General Guidelines" for sample preparation procedures.

Deplete or Positively Isolate CD25⁺ T Cells

The protocol is based on 1 mL (2.5×10^7) MNC or 1 mL whole blood/buffy coat as starting sample, but is scalable from $2.5 \times 10^7 - 5 \times 10^8$ (1–50 mL). When working with lower volumes than 1 mL, use the same volumes as indicated for 1 mL. When working with larger volumes, scale up all volumes accordingly, as shown in Table 1.

- 1. Transfer 1 mL cells (2.5×10^7) to a tube and add 25 μ L pre-washed and re-suspended beads.
- 2. Incubate for 20 min (positive isolation) or 30 min (depletion) at 2°C to 8°C with gentle tilting and rotation.
- 3. Place the tube in a magnet for 2 min.
- For depletion; transfer supernatant to a new tube for further use and discard the beads.

or

For *positive isolation*; while the tube is still in the magnet, carefully remove and discard the supernatant.

- 5. Remove the tube from the magnet and add 1 mL Isolation Buffer, pipet 2–3 times (or vortex 2–3 sec) and place the tube in a magnet for 2 min. While the tube is still in the magnet, carefully remove and discard the supernatant.
- 6. Repeat step 5 at least once to wash the bead-bound CD25⁺ T cells. This step is critical to obtain a high purity of isolated cells.
- 7. Resuspend the cell pellet in preferred cell medium.

Keep the cells on 2°C to 8°C until further use in downstream applications.

Table 1: Volumes for isolation/depletion of human CD25 $^{+}$ T cells. This protocol is scalable from 2.5×10^{7} to 5×10^{8} cells.

Step	Step description	Small scale (1X)	Large scale (10X)
	Recommended tube size	5 mL	15 mL
	Recommended magnet	DynaMag [™] -5	DynaMag [™] -15
1*	Sample volume (MNC/blood/buffy)	1 mL	10 mL
1**	Bead volume	25 μL	250 μL
5-6	For positive isolation only: Wash cells (Isolation Buffer)	3 × ~1 mL	3 × ~10 mL

^{* 2.5 × 107} MNC/mL.

Description of Materials

Dynabeads® CD25 are uniform, superparamagnetic polystyrene beads (4.5 µm diameter) coated with a primary monoclonal mouse IgM antibody specific for the CD25 membrane antigen, which is predominantly expressed on human activated T cells, activated B cells and regulatory T cells.

Related Products

Product	Cat. no.
DynaMag [™] -5	12303D
DynaMag [™] -15	12301D
DynaMag [™] -50	12302D
HulaMixer® Sample Mixer	15920D
Dynabeads® Regulatory CD4+CD25+ T Cell Kit	11363D
Dynabeads® FlowComp™ Flexi	11061D

REF on labels is the symbol for catalog number.

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^{**} If very high cell-depletion efficiency is required, increase the beads volume up to double the recommended amount.