

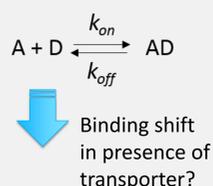
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ABSTRACT

Despite the significant progress made in the drug transporter field, predicting clinical Drug-Drug Interactions (DDIs) based on *in vitro* transporter studies still remains a major challenge. Specifically, the extent of DDIs in terms of changes in C_{max} and AUC of victim drugs is often significantly underestimated using *in vitro* transporter inhibition constants (K_i/IC₅₀), as exemplified by the well-studied OATP mediated DDIs between statins and rifampicin, and the recently reported dolutegravir-metformin interaction. Using OATP1B1 and OCT2 as examples, this talk will discuss the effect of protein binding on "in vivo" transporter inhibition and present a "binding equilibrium shift" hypothesis to explain our experimental findings. Dose-dependent OCT2 and OATP1B1 inhibitions by various drugs with both high and low plasma protein binding, were assessed in protein-free HBSS and protein-rich solutions including HBSS with 4% albumin, 100% human serum and 100% bovine serum. Strikingly, for highly protein bound drugs, unbound fraction (f_u) adjusted IC₅₀ values determined in protein-rich solutions were significantly lower than that obtained from assays using protein-free HBSS. For example, dolutegravir, a HIV integrase inhibitor with >99% protein binding in the human, was significantly more potent in inhibiting OCT2 mediated metformin transport in serum than in HBSS, with f_u adjusted IC₅₀ values of 87nM and 536nM, respectively. In addition, the f_u adjusted permeability of dolutegravir tested in serum was also >5x higher than that assessed in HBSS. On the contrary, the f_u adjusted IC₅₀ for low protein bound Cimetidine (in vivo protein binding ~20%) was independent on assay matrix. These data suggest that for highly protein bound drugs, the "actual" free drug concentrations in presence of transporters and other drug binding membrane proteins could be substantially higher than the calculated free concentration based on f_u measured *in vitro*, as high affinity binding to cell membrane proteins could effectively change the equilibrium of nonspecific binding between drug and serum proteins. Lastly, the talk will present a partial PBPK model incorporating total plasma drug concentration and *in vitro* transporter IC₅₀s obtained with human serum, demonstrating improved IVIVE of dolutegravir and rifampicin associated DDIs. These studies, for the first time to our best knowledge, showed that for highly protein bound drugs, conventional approach based on unbound drug concentration may lead to underestimation of *in vivo* transporter inhibition. As such, we may consider conducting *in vitro* transporter studies in human serum and using total plasma drug concentration for modeling and prediction of transporter mediated DDIs.

BACKGROUND ON PROTEIN BINDING

- At binding equilibrium between Albumin (A) and Drug (D) *in vitro*, the binding dissociation constant $K_d = \frac{k_{off}}{k_{on}} \cdot \frac{[D]}{[AD] + [D]}$
- Unbound fraction at equilibrium is given, $f_u = \frac{[D]}{[AD] + [D]}$
- However, in the presence of additional disposition mechanism, is f_u measured under equilibrium condition appropriate for estimating the effective unbound drug concentration *in vivo*?



- Traditional way of *in vitro* – *in vivo* extrapolation (IVIVE) and DDI prediction goes
 1. Measure IC₅₀ in protein free assay buffer (e.g. HBSS or Krebs-Henseleit buffer)
 2. Measure drug f_u at equilibrium *in vitro* (e.g. rapid equilibrium dialysis)
 3. Use f_u to adjust plasma concentration *in vivo* (C_{total} × f_u) to predict substrate clearance and DDI potentials for perpetrators (e.g. $R = 1 + f_u \cdot [C_{max}]/K_i$)

- Nevertheless, this process required drug-dependent scaling factors for uptake transporters to correlate the observed drug clearance (Jones, H. et al, 2012 DMD). Particularly, Duan, P. et al. reported that such approach under-predicted OATP1B1 inhibition potency of certain perpetrators, e.g. rifampicin-pitavastatin DDI.

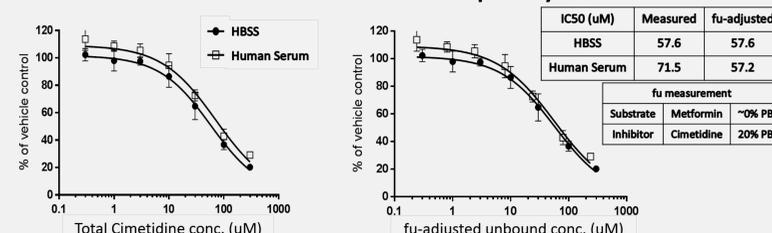
Pitavastatin – Rifampicin DDI Prediction on AUCR (95% CI) (Modified from P., Duan, P. Zhang and L. Zhang, US FDA, ASCPT 2015, PT-17)		
Predicted AUCR	Predicted AUCR with adjusted OATP1B1 Ki (10 fold less)	Observed AUCR
1.89 (1.69-1.98)	5.96 (4.69-6.01)	6.74 (4.73-8.74)

METHODS: ASSESSING TRANSPORTER INHIBITION IN HBSS AND HUMAN SERUM

1. MDCK cells were seeded at 60K cells/well and cultured on PCF porous membrane inserts (0.4 μm pore size, Millipore) using standard culture conditions.
2. Cells were transiently transfected by proprietary Opti-Expression™ using cDNAs encoding OATP1B1 or OCT2 alone, or five transporters shown below.
3. Radioisotope-labeled transporter substrates were applied to the basal side of the cell membrane for uptake assays in presence of various inhibitor concentrations. Actual inhibitor concentrations were measured with LC/MS.
4. Transport assays were conducted in HBSS only or in 100% human serum (HS).

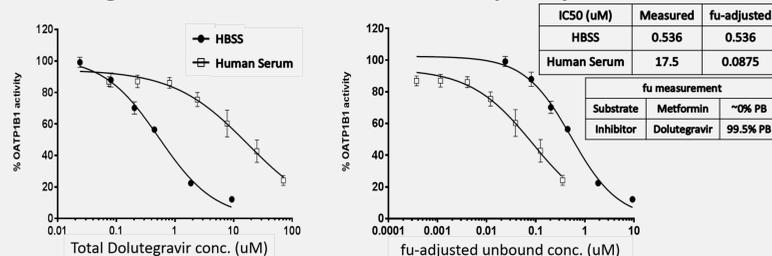
RESULT 1: PROTEIN BINDING EFFECT ON IC50

1. Cimetidine inhibition on Metformin transport by OCT2



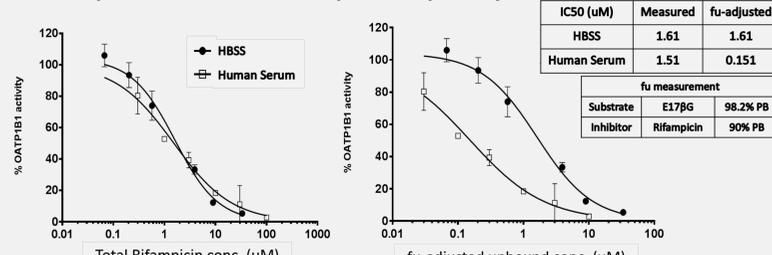
- IC₅₀ of the low protein binding Cimetidine was not affected by serum protein after f_u adjustment.

2. Dolutegravir inhibition on Metformin transport by OCT2



- After f_u adjustment, the inhibition potency of the high protein binding Dolutegravir in human serum was 6.2x higher than that assessed in HBSS

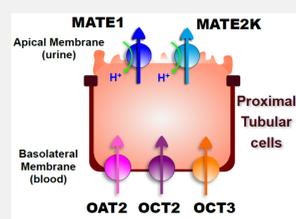
3. Rifampicin inhibition on E17βG transport by OATP1B1



- After f_u adjustment, the inhibition potency of the high protein binding Rifampicin in human serum was 10x higher than that assessed in HBSS

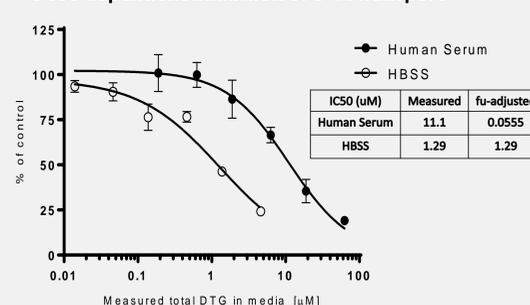
RESULT 2: DOLUTEGRAVIR IC50 MEASUREMENT FOR IVIVE (QUINTUPLE MODEL)

- MDCK-II monolayers expressing a combination of five transporters known as "Quintuple model" with basal OCT2, OAT2, OCT3 and apical MATE1 and MATE2K.
- Radiolabeled Metformin was measured after B>A transcellular transport.



"Quintuple Model" – Zhang Y. et al., 2015 DMD

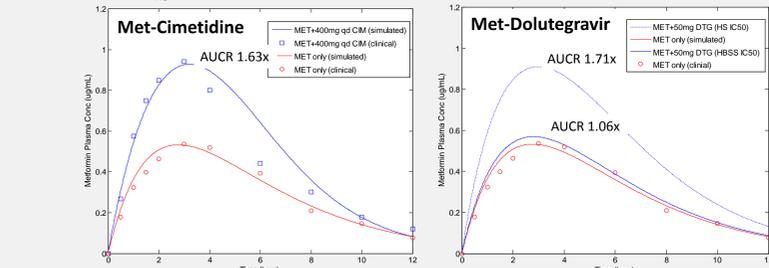
Dose dependent inhibition of B>A transport



RESULT 3: TRANSPORTER DDI MODELING

1. Metformin-Dolutegravir DDI as result of renal OCT/MATE inhibition

- Metformin 100% renally cleared, fitted to PK data from literature¹
- CL_{ns} (net secretion clearance for control), is modified in presence of an inhibitor with [I] and K_i: $CL_{ns}(+I) = \frac{CL_{ns}}{1 + \frac{[I]}{K_i}}$
- Inhibitor-independent *in vitro* to *in vivo* inhibition scaling factor was extrapolated from the metformin-cimetidine DDI data (Left)¹
- Impact of DTG IC₅₀ measured in HS or HBSS was shown (Right)



Predicted MET PK change	MET alone (fitted)	CIM 400mg (Single dose)		IVIVE of DTG 50mg qd (Single dose)	
		in vitro assay in human serum	Using IC ₅₀ from human serum	Using IC ₅₀ from HBSS	Using IC ₅₀ from HBSS
C _{max} Ratio	0.94	1.63 vs. 1.65 ¹	1.60 vs. 1.66 ²	0.99 vs. 1.66 ²	
AUC ₍₀₋₁₂₎ Ratio	1.0	1.63 vs. 1.47 ¹	1.71 vs. 1.79 ²	1.06 vs. 1.79 ²	

2. Pitavastatin-Rifampicin DDI as result of hepatic OATP1B1 inhibition

- Rifampicin PK model: minimal PBPK with gut absorption, renal and hepatic clearances fitted to two sets of PK data in literature³
- Pitavastatin PK model: modified from extended clearance concept model with 5 extrahepatic compartment (EHC) model⁴ with hepatic elimination
- Simultaneous simulation with only OATP1B1 mediated uptake clearance (PS_{inf}) is inhibited by plasma rifampicin concentration. Two IC₅₀s obtained from HBSS or 100% human serum were compared.

P.O admin (FaFg=1)	Pitavastatin 2mg only	Pitavastatin 2mg with 600 mg Rifampicin	
Simulated K _i	No inhibitor	K _i = 16 uM (from IC ₅₀ in HBSS)	K _i = 1.5 uM (from IC ₅₀ in human serum)
C _{max} Ratio	28.0	46.3 (1.7x)	124.2 (4.4x)
AUC Ratio	41.5	70.7 (1.7x)	319.7 (7.7x)

CONCLUSION

- For highly protein bound inhibitors, Dolutegravir and Rifampicin, the apparent IC₅₀s measured in 100% human serum were significantly lower than what were calculated based on f_u and the intrinsic transporter IC₅₀s assessed in HBSS. For low protein binding Cimetidine, such difference was not observed.
- When IVIVE model was simulated with IC₅₀ measured in human serum, Dolutegravir appeared to be a clinically relevant inhibitor of renal secretion of metformin, which is mediated primarily by OCT2 and MATEs.
- Pitavastatin-Rifampicin associated DDI was predicted to be similar to the previously observed AUCR data (background) only when applied with IC₅₀ measured in human serum.
- For highly protein bound inhibitors, it may be useful to obtain their apparent IC₅₀s by running *in vitro* assays in 100% human serum to predict their DDI potentials, e.g., given as ratio R in the presence of an inhibitor.

$$R = 1 + \frac{f_u \cdot I_{max}}{K_{i-HBSS}} \rightarrow R = 1 + \frac{f_u \cdot I_{max}}{f_u \cdot K_{i-Human\ Serum}} = 1 + \frac{I_{max}}{K_{i-Human\ Serum}}$$

Reference

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4. Watanabe T. et al., J Pharmacol Exp Ther., 2009 Feb;328(2):652

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